Fecal Cryptosporidium parvum Antigen ELISA Kit

Catalog Number:
CRY35-K01 (1 x 96 wells)

For Research Use Only. Not for use in diagnostic procedures.

v 1.0
INTENDED USE
This Eagle Biosciences microplate-based ELISA (enzyme linked immunosorbent assay) kit is intended for the qualitative detection of Cryptosporidium parvum antigen in feces. The Eagle Biosciences Cryptosporidium parvum Antigen ELISA Assay Kit is for Research Use Only and is not intended for diagnostic or therapeutic purposes.

SUMMARY OF PHYSIOLOGY
Cryptosporidiosis is one of the main causes of persistent diarrhea in the developed world. It is caused by the presence of Cryptosporidium parvum oocysts in the gastro-intestinal tract. This parasite is known to be highly pathogenic and its infectious stage is transmitted by faecal-oral contract. It is also an opportunistic pathogen found in immunocompromised patients. The symptoms of cryptosporidiosis are watery diarrhea, stomach cramps, weight loss, nausea, and fever. In industrialized countries, 2-2.5% of diarrheal hospitalized patients shed C. parvum oocysts. Ten percent of AIDS patients have chronic cryptosporidiosis and this figure can be as high as 40% in certain developing countries. C. parvum is diagnosed by either Ziehl-Neelsen stain or immunofluorescence in smears of unconcentrated specimens.

ASSAY PRINCIPLE
The Eagle Biosciences Cryptosporidium parvum Antigen is a “sandwich” ELISA designed, developed and produced for the qualitative measurement of Cryptosporidium parvum antigen in stool specimen. The assay utilizes the microplate-based enzyme immunoassay technique by coating highly purified antibody onto the wall of microtiter well. Assay controls and fecal specimen are added to microtiter wells of microplate that was coated with a highly purified polyclonal anti-Cryptosporidium parvum antibody on its wall. The Cryptosporidium parvum antigen will be bound to the antibody coated plate after an incubation period. The unbound matrices are washed away and a HRP-conjugated monoclonal antibody which specifically recognizes the protein of Cryptosporidium parvum is added for further immunoreactions. After an incubation period, an immunocomplex of “Anti-Cryptosporidium Antibody – Cryptosporidium parvum Antigen – HRP-conjugated Anti-Cryptosporidium Tracer Antibody” is formed if Cryptosporidium parvum antigen is present in the test sample. The unbound tracer antibody and other protein or buffer matrix are removed in the subsequent washing step. HRP-conjugated tracer antibody bound to the well is then incubated with a substrate solution in a timed reaction and then measured in a spectrophotometric microplate reader. The enzymatic activity of the tracer antibody bound to C. parvum proteins captured on the wall of each microtiter well is directly proportional to the amount of Cryptosporidium parvum antigen level in each test specimen.

REAGENTS: Preparation and Storage
This test kit must be stored at 2 – 8°C upon receipt. For the expiration date of the kit refer to the label on the kit box. All components are stable until this expiration date. Prior to use allow all reagents to come to room temperature. Reagents from different kit lot numbers should not be combined or interchanged.

1. Anti-Cryptosporidium Antibody Coated Microplate
One microplate with 12 x eight strips (96 wells total) coated with highly purified Anti-Cryptosporidium antibody. The plate is framed and sealed in a foil zipper bag with a desiccant. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

2. **Anti-Cryptosporidium Tracer Antibody**
   One vial containing 0.6 mL concentrated horseradish peroxidase (HRP)-conjugated monoclonal Cryptosporidium antibody in a stabilized protein matrix. This reagent must be diluted with Tracer Antibody Diluent before use. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

3. **Tracer Antibody Diluent**
   One vial containing 12 mL ready-to-use buffer. It should be only used for antibody dilution according to the assay procedures. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

4. **ELISA Wash Concentrate**
   One bottle contains 30 mL of 30-fold concentrate. Before use the contents must be diluted with 870 mL of demineralized water and mixed well. Upon dilution, this yields a working wash solution containing a surfactant in phosphate-buffered saline with a non-azide, non-mercury preservative. The diluted wash solution may be stored at room temperature and is stable until the expiration date on the kit box.

5. **ELISA HRP Substrate**
   One bottle contains 12 mL of tetramethylbenzidine (TMB) with hydrogen peroxide. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

6. **ELISA Stop Solution**
   One bottle contains 12 mL of 0.5 M sulfuric acid. This reagent should be stored at 2 – 8°C or room temperature and is stable until the expiration date on the kit box.

7. **Cryptosporidium Antigen Controls**
   One vial contains Cryptosporidium negative control and another vial contains inactivated Cryptosporidium positive control (30470). Both controls are in a liquid bovine serum albumin-based matrix with a non-azide preservative. The positive control is a dilution of highly purified Cryptosporidium parvum oocysts. After the first use, the controls should be stored at -20°C or below for long-term storage.

8. **Concentrated Patient Sample Diluent**
   One bottle contains 30 mL of 20-fold concentrated buffer matrix with protein stabilizers and preservative. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box. Before use the concentrated buffer must be diluted with 570 mL of distilled water and mixed well. Upon dilution, this yields a working patient sample diluent containing a surfactant in phosphate-buffered saline with a non-azide preservative. The diluted reagent can be stored at room temperature and is stable for 8 weeks. It can also be stored at 2 – 8°C and is stable until the expiration date on the kit box.

**SAFETY PRECAUTIONS**
The reagents must be used in a laboratory and are for research use only. Source material for reagents containing bovine serum albumin was derived in the contiguous 48 United States. It was obtained only from healthy donor animals maintained under veterinary supervision and
found free of contagious diseases. Wear gloves while performing this assay and handle these reagents as if they are potentially infectious. Avoid contact with reagents containing TMB, hydrogen peroxide, or sulfuric acid. TMB may cause irritation to skin and mucous membranes and cause an allergic skin reaction. TMB is a suspected carcinogen. Sulfuric acid may cause severe irritation on contact with skin. Do not get in eyes, on skin, or on clothing. Do not ingest or inhale fumes. On contact, flush with copious amounts of water for at least 15 minutes. Use Good Laboratory Practices.

MATERIALS REQUIRED BUT NOT PROVIDED
1. Precision single channel pipettes capable of delivering 10 µL, 50 µL, 100 µL, and 1000 µL
2. Repeating dispenser suitable for delivering 100 µL.
3. Disposable pipette tips suitable for above volume dispensing.
4. Disposable 12 x 75 mm or 13 x 100 glass or plastic tubes.
5. Disposable plastic 1000 mL bottle with cap.
6. Aluminum foil.
7. Deionized or distilled water.
8. Plastic microtiter well cover or polyethylene film.
9. ELISA multichannel wash bottle or automatic (semi-automatic) washing system.
10. Spectrophotometric microplate reader capable of reading absorbance at 450 nm.

SPECIMEN COLLECTION & STORAGE
Fresh fecal sample should be collected by using a plastic sampling device. The collected fecal sample must be transported, kept at 2-8°C and tested within 2 days. A non-preserved sample must be stored below -20°C for a longer storage period.

ASSAY PROCEDURE
1. Reagent Preparation
Prior to use allow all reagents to come to room temperature. Reagents from different kit lot numbers should not be combined or interchanged.

2. Patient Sample Preparation
   (1) Label a test tube (12x75 mm) or a 1.5 ml plastic vial.
   (2) Add 1 mL of Diluted Sample Diluent to each tube or vial.
   (3) Add 100 µL of liquid stool sample to the above tube.
   (4) With solid stool sample, take an equivalent amount (about 50 – 100 mg) with a spatula or a disposable inoculation loop. Suspend the solid stool sample with 1 mL patient sample diluent and mix well on a vortex mixer.
   (5) Centrifuge the diluted fecal sample at 3000 rpm (1500 g) for 10 – 15 minutes. The supernatant can be directly used in the assay. As an alternative to centrifuging, let the diluted samples sit and sediment for 15 minutes and take the clear supernatant for testing.

   Note: If the test procedure is performed on an automated ELISA system, the supernatant must be particle-free by centrifuging the sample.

3. Assay Procedure
   (1) Place a sufficient number of Anti-Cryptosporidium antibody coated microwell strips in a frame to run Cryptosporidium controls and unknown samples in duplicate.
(2) Test Configuration

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<th>ROW</th>
<th>STRIP 1</th>
<th>STRIP 2</th>
<th>STRIP 3</th>
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<tr>
<td>A</td>
<td>Control Negative</td>
<td>SAMPLE 3</td>
<td>SAMPLE 7</td>
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<tr>
<td>B</td>
<td>Control Negative</td>
<td>SAMPLE 3</td>
<td>SAMPLE 7</td>
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<td>C</td>
<td>Control Positive</td>
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(3) Add **100 µL** of controls and diluted patient stool samples into each designated microwell.

(4) Cover the plate with a plate sealer and also with aluminum foil to avoid exposure to light.

(5) Incubate plate at room temperature for **1 hour**.

(6) Prepare working Anti-Cryptosporidium tracer antibody working solution by **1:21 fold** dilution of the Anti-Cryptosporidium Tracer Antibody with the Tracer Antibody Diluent. For each strip, it is required to mix 1 mL of Tracer Antibody Diluent with 50 µL of Tracer Antibody in a clean test tube.

(7) Remove the plate sealer. Decant the contents of each well. Wash each well 5 times by dispensing 350 µL to 400 µL of **diluted wash buffer** into each well and then completely aspirating the contents. Alternatively, an automated microplate washer can be used.

(8) Add **100 µL** of above diluted tracer antibody working solution to each of the wells.

(9) Cover the plate with one plate sealer and also with aluminum foil to avoid exposure to light.

(10) Incubate plate at room temperature for **40 minutes**.

(11) Remove the plate sealer. Decant the contents of each well. Wash each well 5 times by dispensing 350 µL to 400 µL of **diluted wash buffer** into each well and then completely aspirating the contents. Alternatively, an automated microplate washer can be used.

(12) Add **100 µL** of ELISA HRP Substrate into each of the wells.

(13) Cover the plate with aluminum foil to avoid exposure to light.

(14) Incubate plate at room temperature for 15 minutes.

(15) Remove the aluminum foil. Add **100 µL** of ELISA Stop Solution into each of the wells. Mix gently.

(16) Read the absorbance at 450 nm within 10 minutes in a microplate reader.
PROCEDURAL NOTES

1. It is recommended that all controls and unknown samples be assayed in duplicate. The average absorbance reading of each duplicate should be used for data reduction and the calculation of results.
2. Keep light-sensitive reagents in the original amber bottles.
3. Store any unused antibody coated strips in the foil zipper bag with desiccant to protect from moisture.
4. Careful technique and use of properly calibrated pipetting devices are necessary to ensure reproducibility of the test.
5. Incubation times or temperatures other than those stated in this insert may affect the results.
6. All reagents should be mixed gently and thoroughly prior use. Avoid foaming.

INTERPRETATION OF RESULTS

ELISA Reader:

1. Calculate the average absorbance for each pair of duplicate test results.
2. Calculate the cut-off:
   The positive cut-off and the negative cut-off are established by using following formula.

   **Positive Cut-Off** = 1.1 x (mean extinction of negative control + 0.10)

   **Negative Cut-Off** = 0.9 x (mean extinction of negative control + 0.10)

3. Interpret test result
   i. **Positive**: patient sample extinction is greater than the Positive Cut-Off.
   ii. **Negative**: patient sample extinction is less than the Negative Cut-Off.
   iii. **Equivocal**: patient sample extinction is between the Positive Cut-Off and the Negative Cut-Off.

4. Assay quality control
   1. Positive control must show an average OD reading greater than 0.500.
   2. Negative control should show an average OD reading less than 0.200.

EXAMPLE DATA AND CALCULATED CUT-OFF

A typical absorbance data and the resulting negative control and positive control are represented. **This absorbance must not be used in lieu of control values run with each assay.**

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<thead>
<tr>
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<th>OD 450 nm</th>
<th>Average OD 450 nm</th>
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<tbody>
<tr>
<td><strong>Negative Control</strong></td>
<td>0.088</td>
<td>0.088</td>
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<tr>
<td><strong>Positive Control</strong></td>
<td>1.803</td>
<td>1.772</td>
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LIMITATION OF THE PROCEDURE
1. The results obtained with this Eagle Biosciences Fecal Cryptosporidium parvum antigen test kit serve only as a useful aid to diagnosis. However, the test results should not be interpreted as diagnostic in themselves.
2. Bacterial or fungal contamination of stool specimens or reagents, or cross-contamination between reagents may cause erroneous results.
3. Water deionized with polyester resins may deactivate the horseradish peroxidase enzyme.

QUALITY CONTROL
To assure the validity of the results each assay must include both negative and positive controls. For a valid test, the positive control must have an absorbance of at least 0.5 OD units and the negative control must be less than 0.2 OD units. We also recommend that all assays include the laboratory's own controls in addition to those provided with this kit.

PERFORMANCE CHARACTERISTICS
Sensitivity
The sensitivity of this fecal Cryptosporidium parvum antigen ELISA is about 5 ng/ml of Cryptosporidium parvum antigen as determined by testing a series of dilutions of a highly purified sample of Cryptosporidium parvum antigen with assay buffer and the OD reading is above the positive cut-off.

Reproducibility
The reproducibility of this assay is validated by measuring four samples (two negative and two positive) both in a single assay of 12-replicate determinations and in 6 different assays run on different dates. The results showed a consistent test results interpretation for all the samples.

Specificity
The assay does not cross react to following organisms: Giardia, Rotavirus, and Adenovirus.

REFERENCES
Warranty Information

Eagle Biosciences, Inc. warrants its Product(s) to operate or perform substantially in conformance with its specifications, as set forth in the accompanying package insert. This warranty is expressly limited to the refund of the price of any defective Product or the replacement of any defective Product with new Product. This warranty applies only when the Buyer gives written notice to the Eagle Biosciences within the expiration period of the Product(s) by the Buyer. In addition, Eagle Biosciences has no obligation to replace Product(s) as result of a) Buyer negligence, fault, or misuse, b) improper use, c) improper storage and handling, d) intentional damage, or e) event of force majeure, acts of God, or accident.

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For further information about this kit, its application or the procedures in this kit, please contact the Technical Service Team at Eagle Biosciences, Inc. at info@eaglebio.com or at 866-411-8023.